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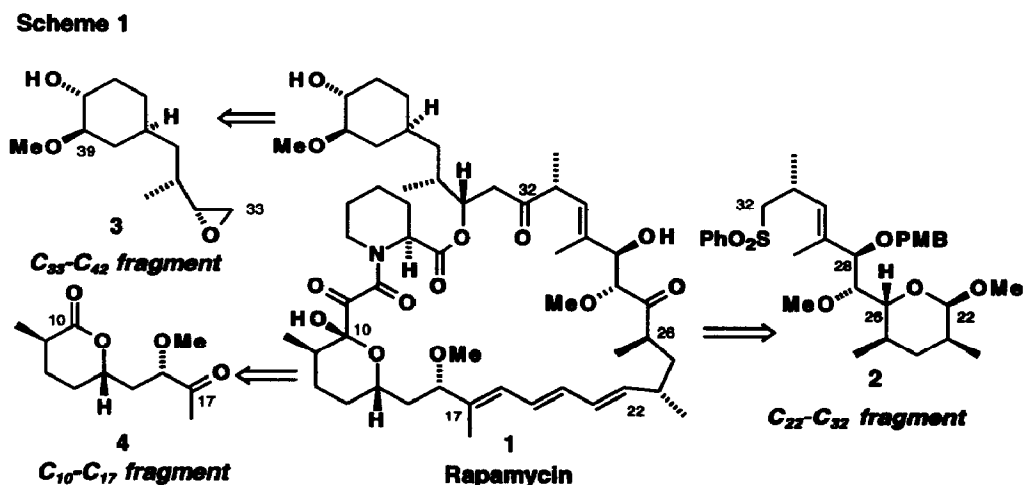
Studies Towards the Total Synthesis of Rapamycin: Preparation of the C₁₀-C₁₇ Carbon Unit.

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Abstract: The preparation of the C₁₀-C₁₇ carbon unit of the immunosuppressive agent rapamycin is reported via two routes leading to the substituted lactone 4.

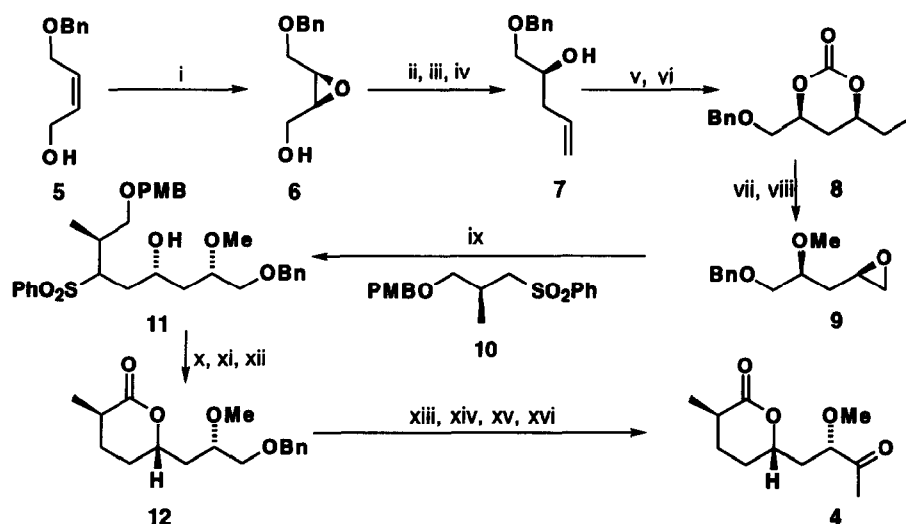
In the preceding papers we have delineated our synthetic strategy towards the immunosuppressive agent rapamycin 1.¹ We also reported on the preparation of key coupling components 2 and 3 for this synthesis and here we describe the construction of the lactone component 4 which constitutes the C₁₀-C₁₇ carbon portion of rapamycin (Scheme 1).



We have devised two routes to the synthesis of 4 which employ a common allylic alcohol 5 as starting material (Scheme 2).² In the first of these syntheses 5 was subjected to an asymmetric Sharpless epoxidation to give 6 in 75% yield with an enantiomeric excess of 92%.^{3,4} Epoxide 6 was then readily transformed to the alkene 7 by pyridine-sulfur trioxide activated dimethylsulfoxide oxidation,⁵ Wittig methylenation and selective opening of the epoxide⁶ in 41% overall yield. Compound 7 was converted to the cyclic carbonate 8 using reaction conditions described by Smith.⁷ The stereoselectivity obtained was generally very good (diastereoselectivity typically of 11:1), however the yields tended to vary between 30 and 57%. Nevertheless,

with good supplies of **8** in hand this was converted to the terminal epoxide **9** by reaction with potassium carbonate in methanol followed by Purdie methylation⁸ with methyl iodide and silver oxide in 95% yield. Epoxide **9** was then coupled with the lithio anion generated from the sulfone **10**⁹ to give **11** in excellent yield employing $\text{BF}_3 \cdot \text{Et}_2\text{O}$ to promote epoxide ring opening.¹⁰ This product was converted to the lactone **12**¹¹ in a sequence of reactions involving desulfonation by lithium naphthalenide reduction, deprotection of the *paramethoxybenzyl* group and finally selective oxidation with tetra-*n*-propylammonium perruthenate (TPAP)¹² to afford **12** in 83% yield.¹³ Further elaboration of the side chain of **12** involved hydrogenolysis of the benzyl group, oxidation with the Dess-Martin periodinane reagent,¹⁴ methyl Grignard addition and a second oxidation which afforded the key lactone coupling partner **4** in 70% yield over the four steps (Scheme 2).¹⁵ This synthetic lactone **4** was identical to a sample previously obtained from the degradation of rapamycin.¹⁶

Scheme 2

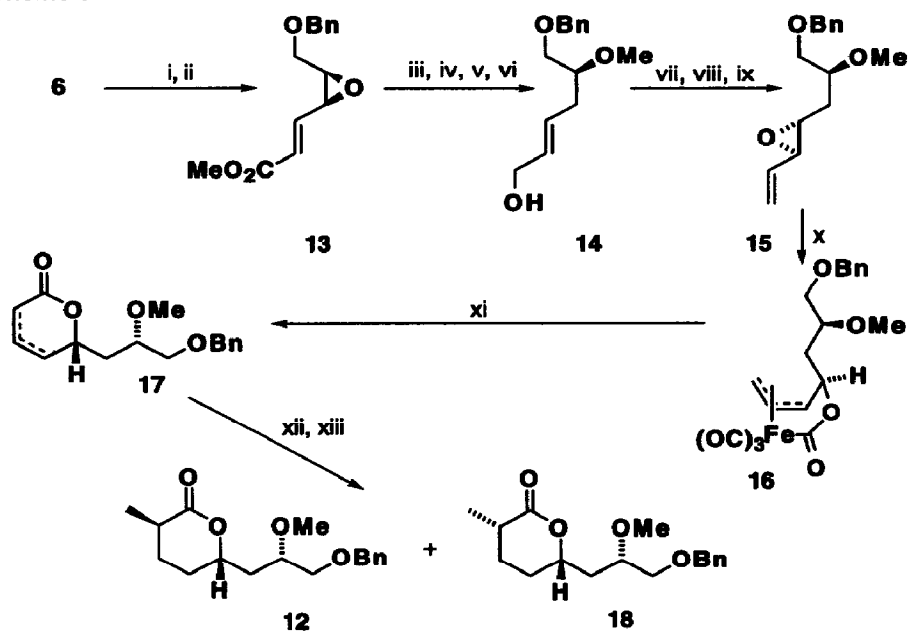


i) $\text{Ti}(\text{O}^i\text{Pr})_4$, (+)-diethyl tartrate, $^t\text{BuOOH}$, CH_2Cl_2 , -25°C , 75%; ii) $\text{py} \cdot \text{SO}_3$, Et_3N , DMSO , CH_2Cl_2 , 0°C to rt; iii) MePPh_3Br , KHMDS , THF , 0°C to rt; iv) DIBAL-H , toluene, -78°C , 41% overall; v) $^n\text{BuLi}$, Et_2O , rt, followed by BOC-ON , THF , rt, 90%; vi) IBr , toluene, CH_2Cl_2 , -85°C , 30-57%; vii) K_2CO_3 , MeOH , rt, 90%; viii) MeI , Ag_2O , DMF , rt, 95%; ix) $^n\text{BuLi}$, THF , -78°C , followed by **9**, $\text{BF}_3 \cdot \text{Et}_2\text{O}$, THF , -78°C to rt, quant.; x) 1M Lithium naphthalenide, THF , -90°C , 72-93%; xi) DDQ , CH_2Cl_2 , H_2O , rt, quant.; xii) TPAP, NMO , CH_2Cl_2 , 4Å powdered molecular sieves, 83%; xiii) $\text{Pd}(\text{OH})_2/\text{C}$, H_2 , 1 atm., EtOAc , rt, quant.; xiv) Dess-Martin periodinane, $^t\text{BuOH}$, py , CH_2Cl_2 ; xv) MeMgBr , Et_2O , THF , -78°C ; xvi) Dess Martin periodinane, $^t\text{BuOH}$, py , CH_2Cl_2 , 70% overall.

In a conceptually different approach¹⁷ to the same lactone **12** we have explored the use of π -allyltricarbonyliron lactone complexes as precursors as these have proved useful to us in several other natural product syntheses.¹⁸ The previously synthesised epoxide **6** was therefore oxidised and treated with methyl diethyl phosphonoacetate to give **13** with excellent *E* selectivity (Scheme 3). Following DIBAL reduction and conversion to the allylic alcohol **14** this was subjected to a catalytic Sharpless epoxidation. Oxidation using TPAP and a Wittig reaction afforded the alkenyl epoxide **15**,¹⁹ in 90% diastereomeric excess as the precursor for the iron carbonyl chemistry. Reaction of **15** with $\text{Fe}_2(\text{CO})_9$ in THF ²⁰ gave the *endo* complex

16²¹ as the predominant product in 72% yield. Exhaustive carbonylation²² with CO at 280 atm in benzene at 70°C gave **17** in 85% yield as a mixture of α,β - and β,γ -unsaturated lactones in a ratio of 7:3 respectively. Hydrogenation employing Adam's catalyst and methylation afforded the required lactone **12** which was identical to the sample prepared previously, together with the epimeric lactone **18**.²³ At present this methylation, using lithium diisopropylamide as base and quenching with methyl iodide, gives **18** and **12** in a 60:40 ratio.²⁴ However it is possible to recycle **18** *via* deprotonation/reprotonation to provide further supplies of **12**.

Scheme 3



i) $\text{Py}\cdot\text{SO}_3$, Et_3N , DMSO, CH_2Cl_2 , 0°C to rt, 80%; ii) LiCl, $(\text{EtO})_2\text{P}(\text{O})\text{CH}_2\text{CO}_2\text{CH}_3$, DBU, CH_3CN , rt, 68%; iii) DIBAL-H, CH_2Cl_2 , -78°C; iv) PivCl , py, CH_2Cl_2 , 0°C, 69% overall; v) NaH, MeI, 0°C, 94%; vi) DIBAL-H, CH_2Cl_2 , -78°C, 88% vii) 10% $\text{Ti}(\text{OPr})_4$, 14% (-)-diethyl tartrate, $^t\text{BuOOH}$, 4Å powdered molecular sieves CH_2Cl_2 , -23°C, 80%; viii) TPAP, NMO, CH_2Cl_2 , CH_3CN , 4Å powdered molecular sieves, 60%; ix) MePPh_3Br , KHMDS, THF, 0°C to rt, 83%; x) $\text{Fe}_2(\text{CO})_9$, degassed THF, 72%; xi) CO, 280 atm., benzene, 70°C, 2 days, 85%; xii) PtO_2 , H_2 , 1 atm., EtOAc , rt, 82% xiii) LDA, THF, -78°C, MeI, 80%.

In summary, we have devised two routes to the lactone **4** which we intend using as the C₁₀-C₁₇ carbon unit during our synthesis of rapamycin.

Couplings of the fragments which have been proposed here and in the preceding papers will be reported at a later date.

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References and footnotes

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2. *cis*-4-Benzoyloxy-2-buten-1-ol, **5** was purchased from Aldrich Chemical Company, Inc. and used without purification.
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11. Data for **12**: $[\alpha]_D^{25} = +16.76$ ($c = 0.72$, CHCl_3); IR (CHCl_3): ν_{max} (cm^{-1}): 2932, 2873, 1728, 1495, 1454, 1376, 1361, 1241, 1177, 1092, 1035, 749, 699; $^1\text{H NMR}$ (250 MHz, CDCl_3 , rapamycin numbering): δ (ppm): 7.31 (5H, m, Ar-H), 4.53 (2H, 2d, $J = 12$ Hz, $\text{CH}_2\text{-Ar}$), 4.32 (1H, m, $\text{C}_{14}\text{-H}$), 3.55 (3H, m, $\text{C}_{16}\text{-H}$ and $\text{C}_{16}\text{-H} \times 2$), 3.37 (3H, s, OMe), 2.39 (1H, m, $\text{C}_{11}\text{-H}$), 1.90 (4H, m, $\text{C}_{12}\text{-H} \times 2$ and $\text{C}_{13}\text{-H} \times 2$), 1.57 (2H, m, $\text{C}_{15}\text{-H} \times 2$), 1.19 (3H, d, $J = 7$ Hz, $\text{C}_{11}\text{-Me}$). $^{13}\text{C NMR}$ (50 MHz, CDCl_3): δ (ppm): 174.2 (C=O), 138.2, 128.4, 127.8 and 127.7 (Ar), 78.9 (C_{16}), 76.4 ($\text{CH}_2\text{-Ar}$), 73.3 (C_{17}), 70.5 (C_{14}), 57.2 (OMe), 37.7 (C_{15}), 36.1 (C_{11}), 29.3 (C_{13}), 28.5 (C_{12}), 17.4 ($\text{C}_{11}\text{-Me}$); mass (EI): m/z : 292 (M^+), 246, 204, 171, 160, 149, 113, 91, 85; HRMS, calculated for $\text{C}_{17}\text{H}_{24}\text{O}_4$: calcd: 292.1674; found: 292.1670.
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15. Data for **4**: $^1\text{H NMR}$, δ ppm (400 MHz, CDCl_3): 1.28 (3H, d, J 7Hz), 1.59 (2H, m), 1.92 (2H, m), 2.05 (2H, m), 2.21 (3H, s), 2.42 (1H, m), 3.38 (3H, s), 3.76 (1H, t, J 6Hz), 4.44 (1H, m).
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17. We thank Dr. Roberto D' Alessio for initial studies in this area.
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19. The diastereomeric excess of **15** was determined by GC analysis.
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21. Data for complex **16**: $^1\text{H NMR}$ (400 MHz, C_6D_6): δ (ppm): 7.33 (5H, s), 4.30 (2H, s), 4.19 (1H, dd, J 5, 12Hz), 3.86 (1H, dd, J 5, 8Hz), 3.62 (1H, dt, J 8, 13.2Hz), 3.34 (2H, d, J 1.5Hz), 3.28-3.41 (1H, m), 3.15 (3H, s), 2.77 (1H, dd, J 1.5, 8.4Hz), 2.72 (1H, dd, J 1.5, 13.2Hz), 1.74 (2H, dd, J 5, 7Hz); $^{13}\text{C NMR}$ (50 MHz, C_6D_6): δ (ppm): 208.8, 207.1, 203.8, 200.1, 139.0, 128.3, 127.8, 90.8, 82.3, 78.1, 73.8, 73.4, 71.4, 58.5, 57.0, 38.8.
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23. Data for **18**: $^1\text{H NMR}$ (250 MHz, CDCl_3): δ (ppm): 7.33 (5H, m, Ar-H), 4.55 (2H, 2d, $J = 12$ Hz, $\text{CH}_2\text{-Ar}$), 4.33 (1H, m, $\text{C}_{14}\text{-H}$), 3.55 (3H, m, $\text{C}_{16}\text{-H}$ and $\text{C}_{16}\text{-H} \times 2$), 3.39 (3H, s, OMe), 2.49 (1H, m, $\text{C}_{11}\text{-H}$), 1.91 (4H, m, $\text{C}_{12}\text{-H} \times 2$ and $\text{C}_{13}\text{-H} \times 2$), 1.55 (2H, m, $\text{C}_{15}\text{-H} \times 2$), 1.19 (3H, d, $J = 7$ Hz, $\text{C}_{11}\text{-Me}$). $^{13}\text{C NMR}$ (50 MHz, CDCl_3): δ (ppm): 174.2, 138.1, 128.4, 127.8 and 127.7, 76.3, 74.9, 73.3, 70.5, 57.2, 36.8, 33.0, 26.8, 25.5, 16.0.
24. Lactones **12** and **18** are readily separable by HPLC.

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